

Bacillus cereus (BC) Nucleic Acid Detection Kit (Fluorescent PCR Method)

Product Number: DTK266

Shipping and Storage

1. Store in dark at -20°C, with a shelf life of 12 months.
2. Low temperature transportation cannot exceed 4 days; After opening, store in the dark at -20°C. The expiration date has no impact. Avoid repeated freezing and thawing, freezing and thawing 6 times will not affect the detection effect.

Component

Component	Specification/Volume μ L/Quantity		Main components
	25T	50T	
qPCR premix (containing enzymes)	400 μ L	800 μ L	Tris, KCl, MgCl ₂ , dNTPs, Taq enzymes, etc
Primer probe Bacillus cereus	100 μ L	200 μ L	Primer probe
Positive control Bacillus cereus	50 μ L	50 μ L	Plasmids containing target detection gene fragments
Negative control	50 μ L	50 μ L	Water treated with diethyl carbonate

Description

This reagent kit is designed based on the principle of fluorescence PCR technology, with specific primers and Taqman probes for Bacillus cereus. It is detected by a fluorescence PCR detector to achieve the detection of Bacillus cereus nucleic acid.

Application

This reagent kit is used for qualitative detection of Bacillus cereus nucleic acid and for auxiliary diagnosis and epidemiological monitoring of Bacillus cereus infection.

Applicable instruments

Suitable for fully automatic fluorescence PCR detectors such as ABI 7500, Bio-Rad CFX96, Roche480, etc.

Specimen collection

1. Sample types: feces, vomit, and other samples.
2. Storage conditions: The collected specimens should be sent for testing in a timely manner. Those tested within 24 hours should be stored at 4°C, and those tested beyond 24 hours should be stored at -70°C, avoiding repeated freezing and thawing.

Protocol

1. Reagent Preparation (Reagent Preparation Area)

Melt the components of the reagent kit at 4°C in the dark, mix thoroughly, and centrifuge immediately. Calculate the number of reagents used N (N=number of samples+1 tube of positive control+1 tube of negative control), configure the reaction system mix according to the table below, add it to an appropriate volume of centrifuge tube, mix thoroughly, and centrifuge immediately. Divide it into 20 μ L PCR reaction tubes/plates and transfer it to the sample processing area.

Component	Volume (μ L)
qPCR premix (containing enzymes)	16 μ L
Primer probe Bacillus cereus	4 μ L
Total volume (reaction system mixture)	20 μ L

2. Sample Processing (Sample Processing Area)

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2.1. Nucleic acid extraction:

Select the appropriate nucleic acid extraction kit to extract nucleic acid, and follow the instructions of the corresponding kit for specific operations.

2.2. Sampling:

Add 5µL of processed nucleic acid, negative control, and positive control to the PCR reaction tube/plate that has been mixed with the reaction system mixture, resulting in a final volume of 25µL. Cover the tube tightly or seal it with a membrane, and perform transient centrifugation followed by detection on a fluorescence PCR amplification instrument.

3. Amplification testing (nucleic acid amplification area)

step	Temperature	Time	cycles
Pre denaturation	95°C	5min	1 cycle
Denaturation	95°C	10sec	40 cycles
Annealing/extension/fluorescence detection*	55°C	40sec	

Note: Fluorescence detection at 55°C in step 2, using FAM as the detection channel.

The ABI series fluorescence PCR instrument does not select ROX calibration, and the quenching group is selected as None.

4. Result analysis

According to the analysis of the image, adjust the start and end values (it is recommended to start from 3-15 and end from 5-20, and adjust the amplification curve of the negative control to be flat or below the threshold line). Click on the analysis button and view the results on the report interface.

Quality control standards

1. Negative control: Ct value>38 or not detected.
2. Positive control: The amplification curve is S-shaped and the Ct value is ≤ 30.
3. The above requirements must be met simultaneously for the same experiment, otherwise this experiment will be considered invalid.
4. Each detection target requires a positive and negative control, and the baseline threshold is adjusted for different targets based on their corresponding negative results.

Result interpretation

1. FAM channel detects the universal type of Escherichia coli.
2. Negative: Ct value>38 or not detected.
3. Positive: The amplification curve is S-shaped and the Ct value is ≤ 35.
4. Suspicious: The amplification curve shows an S-shaped pattern, and the Ct value is between 35 and 38, requiring retesting; If the retest results are consistent, the judgment result is positive. If the CT value is greater than 38 or not detected, the judgment result is negative.

Limitations of detection methods

1. Improper sample collection, transportation, and storage, as well as improper transportation, storage, and configuration of reagents, can all affect experimental results and even lead to false negative results.
2. If there is laboratory contamination, reagent contamination, or sample cross contamination, false positive results may occur.

Performance indicators of reagent kit

1. Minimum detection limit: 1×10^3 copies/mL.
2. Linear detection range: $2 \times 10^3 \sim 1 \times 10^8$ copies/mL.
3. Specificity: It can be detected in all specimens of Bacillus cereus and does not cross with other types.



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Note

1. Each stage of PCR operation should be strictly partitioned to avoid cross contamination.
2. The components of the reagent kit should be thoroughly melted and mixed before use, and centrifuged for a few seconds before use.
3. Each component shall not be interchanged with other products or corresponding ingredients of different batch numbers.
4. If the specimen to be tested is not tested in a timely manner, it should be stored at -20°C or -70°C.
5. The processing of samples should strictly follow biosafety regulations.
6. PCR operators should have experience and receive professional training.
7. This kit is only used for scientific research purposes and is not intended for clinical diagnosis.